On page 7, under the heading of Brief Description of the Drawings, please amend the first seven paragraphs to read as indicated below (a "version with markings to show changes made" is presented in Appendix A):

Figure 1 depicts the nucleotide sequence encoding a human secreted B7-4, B7-4S (SEQ ID NO: 1).

Figure 2 depicts the nucleotide sequence encoding a human B7-4, B7-4M (SEQ ID NO: 3).

Figure 3 depicts the amino acid sequence of human B7-4S (SEQ ID NO: 2) and illustrates the signal, IgV, IgC, and hydrophilic tail domains.

Figure 4 depicts the amino acid sequence of human B7-4M (SEQ ID NO: 4) and illustrates the signal, IgV, IgC, and transmembrane and cytoplasmic domains.

Figure 5 depicts the nucleotide sequence of murine B7-4 (SEQ ID NO: 10).

Figure 6 depicts the amino acid sequence of murine B7-4 (SEQ ID NO: 11).

Figure 7 depicts an alignment of the human B7-4M (SEQ ID NO: 4) and murine B7-4 (SEQ ID NO: 11) amino acid sequences. Identical residues are reiterated between the two sequences.

On page 96, please amend the second paragraph to read as indicated below (a "version with markings to show changes made" is presented in Appendix A):

Oligonucleotides with the sequence CAGCTATGGTGGTGCCGACTACAA (SEQ ID NO: 5) and AGGTGCTAGGGGACAGTGTTAGACA (SEQ ID NO: 6) from these ESTs were synthesized. These oligonucleotides were used to prime a PCR reaction using as template cDNA prepared by reverse transcription of mRNAs from the spleen of a case of follicular lymphoma, activated B cells, INF-γ activated keratinocytes, normal spleen, and placenta. Conditions were 94°C, 1 min; 94°C, 30 sec, 56°C, 30 sec, 68°C, 1 min for 35 cycles; 68°C, 3 min, hold 4°C. All templates gave a band of the expected size of 389 bp. The 389 bp product

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from the PCR of INF-γ activated keratinocytes was purified by agarose gel electrophoresis and 0.12 ng was used as a template in a PCR reaction containing 0.05 mM biotin-21-dUTP and the above primers. Conditions were 94°C, 1 min; 94°C, 30 sec, 56°C, 30 sec, 68°C, 2 min for 20 cycles; 68°C, 5 min, hold 4°C. The biotinylated PCR product was purified on a Nucleospin column (Clontech) and used as a probe in the ClonCapture cDNA selection procedure (Clontech). 60 ng of denatured, biotinylated PCR product was incubated with 2 mM $CoCl_2$, 1 X RecA buffer, 1 μg of RecA protein, 1X ATP in a final volume of 30 μl. The reaction was incubated at 37° for 15 min. 0.7 μg of plasmid DNA of an activated keratinocyte cDNA library and 0.4 μg of a human placental cDNA library was added and incubation continued for 20 min. 50 ng of EcoRV digested lambda DNA was added to the reaction and incubated 5 min. 0.6 μl of 10% SDS and 5.6 μg of proteinase K were added and incubated at 37° for 10 min. Proteinase K was inactivated by adding 1 μl of 0.1 M PMSF. Streptavidin magnetic beads were preincubated with 5 μg of sheared salmon sperm DNA for 10 min and the beads

Replacement Paragraph

On page 97, amend paragraph 1, to read as indicated below (a "version with markings to show changes made" is presented in Appendix A):

captured with a magnet, the supernatant removed, and the beads resuspended in 30 μl of binding buffer (1 mM EDTA, 1 M NaCl, 10 mM Tris-HCl, pH 7.5). The beads were added to the reaction and the reaction incubated for 30 min at room temperature with gentle mixing. The beads were captured with a magnet and the supernatant removed. The beads were washed with 1 ml of washing buffer (1 mM EDTA, 2 M NaCl, 10 mM Tris-HCl, pH 7.5), beads were captured with a magnet and the supernatant removed. The wash procedure was repeated 3 times. One ml of sterile H_2O was added to the washed beads, incubated 5 min at 37°, beads were captured on a magnet and the supernatant removed. Captured

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